## TUMOR-PROMOTING COMPOUNDS FROM EUPHORBIA COOPERI

DI- AND TRIESTERS OF 16-HYDROXY-12-DESOXY-PHORBOL.

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(Received in UK 19 December 1969; accepted for publication 16 January 1970)

By a combination of liquid-liquid extraction methods with adsorption chromatography from latex of Euphorbia cooperi<sup>+)</sup> two new irritant and tumor promoting compounds C (I), C' (II) have been isolated.

The mass spectra of C ( $M^+516$ ) and C' ( $M^+558$ ) show that they are di- and triesters respectively of the same parent alcohol (III: mw = 364). Further, the fragmentations  $M^+ - 88$  and  $M^+ - 100$  in the diester C and  $M^+ - 60$ ,  $M^+ - 88$ and  $M^+ - 100$  in the triester C' indicate that in both compounds the estergroups may contain the same  $C_4^-$  (88) and  $C_5^-$ acid with one C=C-double bond (100) respectively in addition to acetic acid (60) in the triester C'. Acid catalysed selective transesterification (HC10<sub>4</sub>/MeOH) of the acetic acid ester group in C' yields C.



\*) We are greatly indebted to Dr.R.Dyer and Mr.G.Vahrmeijer,Department of Agricultural Technical Services, Botanical Research Institute, Pretoria, Republic of South Africa, for supply of latex. By analysis of the nmr-spectrum of I the ester groups in this triester may be further characterized and identified: the sharp singlet at 2,03 ppm is characteristic of an acetyl group, the doublet (J = 7 Hz) at 1,13 ppm of the geminal methylgroups of an isobutyric acid residue  $(C_4$ -acid). The CH<sub>3</sub>-peaks at 1,91 and 1,96 ppm show the third acid residue to be tiglic acid  $(C_5$ -acid with one C=Cdouble bond). The couplings  $(J_1=7 \text{ Hz}, J_2=1 \text{ Hz})$  of these CH<sub>3</sub>-signals with the =CHsignal at 6,07 ppm can be shown by double resonance experiments.

Base catalysed transesterification (NaOCH<sub>3</sub>/MeOH) from I yields a monotigliate IV. Trials to obtain the parent alcohol failed, instead by a rapid reaction IV ( $R_{\rm p}$  o,13, thin layer, Silicagel b/e<sup>+)</sup>=1/3) is converted to a less polar compound ( $R_{\rm p}$  o,37 system as IV). Acetylation of IV with Ac<sub>2</sub>O/pyridine yields a monotigliate-diacetate (V; M<sup>+</sup> 530) with an uv-spectrum  $\sum_{\rm max}^{\rm CH_3OH}$  (245), 330 nm (£5600,70) max very similar to phorbol-12,13,20-triacetate (1). Also the nmr-spectrum of V (chart 1, table 1) shows most of the signals of phorbol-12,13,20-triacetate with similar chemical shifts and multiplicities (1,2). However, the signal of one of the CH<sub>3</sub>-groups at the cyclopropanering (CH<sub>3</sub>-16 or 17) as well as the signal of the -CH(OAc) group (H-12) as exhibited by phorbol-12,13,20-triacetate are not present in V. Furthermore a new signal of a -CH<sub>2</sub>(OAc) group at 4,16 ppm and of a -CH<sub>2</sub> group at 2,1 ppm is observed in V. These results suggest that the parent alcohol of I and II is an isomer of phorbol with OH-12 translocated from position 12 to either one of the methylgroups in position 16 or 17 (see III).



Compound  $R_{\rm F}$  0,37 obtained in trials to obtain III from IV (see above) is identical with crotophorbolon (VI) by melting point, mixed melting point, ir-spectra and  $\alpha \frac{20}{D} = 173^{\circ}$  (1% in EtOH). VI is one of the products of the reaction of phorbol

+) b = benzene, e = ethylacetate

with 0,02 n sulfuric acid (2,3). The transformation of IV to crotophorbolon (VI) may be understood as base catalysed elimination of OH-16 (see scheme). It thus proves the structure of the parent alcohol III as derived from nmr-measurements including the absolute configurations at seven out of eight asymmetric centres at C-4,8,9,Io,I1,I3,I4. Thus III is a 4,9,I3,I6(or 17),20-Pentahydroxy-tigliadien-1,6-on-(3). The absolute configuration at the eighth asymmetric centre (at C-15) may be judged from sterical considerations using Dreiding models. With the -CH\_OCOCH\_ group of V in B-(endo)-position a strong intramolecular H-bridge from the estercarbonyl to OH-4 may be expected. Because the position of the signal of OH-4 (by D<sub>2</sub>O-exchange) in the nmr-spectra of V and of phorbol-12,13,20-triacetate is identical OH-4 does not seem to participate in a H-bridge. Therefore most probably the -CH\_OCOCH\_ group in V and the -CH\_OH-group in III are in a-(exo)-position, i.e. cis-configuration with respect to the reference atom H-14. From the point of view of diterpene biosynthesis too an exo-position of -CH\_OH at C-15 is more likely: usually the straight chain precursor geranyl-geraniol-pyrophosphate is cyclised first and the resulting product subsequently oxygenated. In a corresponding perhydroazulene precursor of phorbol (4) the methylgroup 17 in ß-or endo-position <sup>+)</sup> would not be as easily accessible for hydroxylation as the methylgroup 16 in a-or exo-position. Therefore III is most probably 4,9,13,16,20-Pentahydroxytiglia-dien-1,6-on-(3).

Financial support by the Deutsche Forschungsgemeinschaft is gratefully acknowledged.

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- +) Of both methylgroups in 15-position the  $\beta$ -methyl was defined as number 17 (5).



Chart 1: loo MHz nmr-spectrum of the monotigliate-diacetate (V) in  $CDCl_3$  with tetramethylsilane (d = 0,00 ppm) as internal standard. Ti = signals of the tiglic acid residue.

irradiated	at ppm	observed	at ppm	change in multiplicity	removed coupling (J in Hz)
H-1	7,56	н <sub>3</sub> -19	1,80	dd →→ d	<1
H-lo	3,32	н <sub>3</sub> –19	1,80	$dd \longrightarrow s$ (broad)	2
H-lo	3,32	н-1	7,56	<b>sharpenin</b> g	-
H-7	5,66	н-8	3,06	$t \longrightarrow d$	5,5
H-7	5,66	н <sub>2</sub> -5	2,46	sharpening	
н-8	3,06	н-7	5,66	d → s	5,5
н-8	3,06	н-14	1,25	d → s	5
н-14	1,25	н-8	3,06	$t \longrightarrow d$	5
H-11	2,06	н <sub>3</sub> -18	o,92	d→ s	6

Table 1: Decoupling in the nmr-spectrum of V by double resonance experiments.